



Purgo

Bacterial Persistence

The study below tested how long a surface cleaned or treated with Purgo would remain hygienic.

Summary points include:

1. Purgo was still working after **4 days**.
2. During and until completion of the 4-day test, Purgo eradicated **99.9%** of a variety of strains including Staphylococcus, E-Coli, Listeria and Salmonella among others.

Scroll down to view the complete study*.

*Milouda Labs, Israel

28/6/2020

To– ECOTIV CLEAN**From – Milouda laboratories – Ronit Ben Avraham****Microbiological Test –****Chemical disinfectants and antiseptics - Quantitative carrier test**
For the evaluation of bactericidal and fungicidal activity for
instruments used in the medical and food areas**Laboratory Number: 20055117****Sample description: PURGO –
DISINFECTION SUSPENSION****Date sample received: 1/6/2020****Date Tested: 3/6/2020****1. Standard:**

The test was conducted based on Israeli Standard 1944, BS EN 14561
"Evaluation of bactericidal activity" and AAMI TIR 12 (2010).

Test Purpose:

This test was conducted in order to define the antimicrobial effectiveness of
the disinfectant preparation (PURGO).

Inoculation:

- 1.1 Stainless steel surfaces (4/4cm) were sterilized by steam.
- 1.2 The surfaces were inoculated with the following bacteria (four surfaces
for each microorganism) –

<i>Staphylococcus aureus</i> ATCC 6538
<i>Pseudomonas aeruginosa</i> ATCC 9027
<i>Escherichia coli</i> ATCC 8739
<i>Aspergillus niger</i> ATCC 16404
<i>Enterococcus faecalis</i> ATCC 51299
<i>Lactobacillus plantrarum</i> ATCC 14917
<i>Salmonella typhimurium</i> ATCC 14028
<i>Enterobacter aerogenes</i> ATCC 13048

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Remarks:

1. The laboratory operates under organized working procedures which correlate to the international standard ISO/IEC 17025 in those disciplines where accreditation has been granted.
2. The microbiological tests are within the recognition framework of the Ministry of Health as published in the registrations.
3. The results are related only to the tested sample.
4. This document may be referred to in its entirety, and no part may be quoted or copied to other documents.
5. Sampling was provided by and is the sole responsibility of the customer.
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<i>Saccharomyces cerevisiae</i> ATCC 51299
<i>Listeria monocytogenes</i> ATCC 19115
<i>Bacillus atrophaeus</i> spores ATCC 9372

The bacterial suspensions were diluted using soil (ATS - containing Proteins Healthmark (MI, US)) to give a final concentration of bacteria Of 10^5 - 10^6 per/surface (about 100 μ l from each suspension was added To the soil according to cell turbidity).

- 1.3 One surface was not inoculated – negative control.
- 1.4 The surfaces were left to dry in biohazard hood for 30 minutes.
- 1.5 The surfaces were sprayed with the disinfection suspension and left in the biohazard for **4 days** and then tested.

2. Test Procedure:

- 2.1 Two surfaces before disinfection and cleaning for each microorganism were placed aseptically into sterile cups. 100 ml were added to each sample (Neutralizing solution lot 904) and vortexed for 1 minute and then the diluted sample was plated according to the pour plate technique using warm TSA (lot 949) or SDA (lot 16598) or APT (lot 16584).
- 2.2 The plates were incubated for 72 hours at 30-35°C or 120 hours at 20-25 °C for yeasts and moulds. After incubation of the test plates, the Microorganisms were counted on each plate.
- 2.3 The remaining surfaces (two for each microorganism) were subjected to disinfection according to manufacturer's instructions (Spraying the surfaces with the tested sample). Two surfaces after disinfection were put into cups and 0.1 ml of Neutralizing solution was spread on each surface. The surfaces were then diluted with 10 ml (BPS+1% Tween 80 lot 16358) and vortexed for 1 minute.
- 2.4 Then the eluent was plated in the pour plate technique using TSA or SDA or APT. One surface was touched using rodac plates by (TSA +Lec +Polys 80)
- 2.5 The plates were incubated as defined in 2.1 and then the microbial count was determined per surface.

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3. Results:

Bacteria/Yeast/Mould	Before disinfection CFU/surface	After disinfection 30 seconds CFU/surface
<i>P. aeruginosa</i> ATCC 9027	1800 2600	<10 <1
<i>S.aureus</i> ATCC 6358	102,000 122,000	<10 <1
<i>E.coli</i> ATCC 8739	5600 2800	<10 <1
<i>Aspergillus niger</i> ATCC 16404	46,000 38,000	160 120
<i>Enterococcus faecalis</i> ATCC 51299	13,200 12,800	8600 9400
<i>Lactobacillus plantrarum</i> ATCC 14917	8200 6400	<10 <1
<i>Salmonella typhimurium</i> ATCC 14028	1,860,000 1,920,000	520 440
<i>Enterobacter aerogenes</i> ATCC 13048	13.200 10,400	2400 2200
<i>Saccharomyces cerevisiae</i> ATCC 51299	3400 2600	<10 <1
<i>Listeria monocytogenes</i> ATCC 19115	3600 3200	<10 <1
<i>Candida albicans</i> ATCC 10231	3400 2800	<10 <1
NC	<10	

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4. Conclusion:

According to the test results, the disinfection using **PURGO** in the presence of organic soil was able to reduce at least 3-5 magnitudes (99.9%) for *P.aeruginosa*, *L.plantarum*, *L.monocytogenes*, *S.aureus*, *S.typhimurium*, *C.albicans*, *S.cerevisiae* and *E.coli*.

A.niger, *E.fecalis* and *E.aerogenes* were reduced by two magnitudes.

*****End of Test Results*****

Authorized Signature: _____

Yehudit Ben Avraham PhD
Professional Manager
Microbial Laboratory

Preformed by: _____

צפנתה חיון

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